Relevant Preclinical Models in Recurrent Acute and Chronic Pancreatitis

William Salerno, Chloe Nelson, Lola Rahib

06-12-2024

Introduction

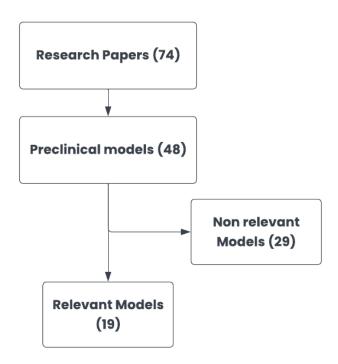
Recurrent acute pancreatitis (RAP) and chronic pancreatitis (CP) lack effective disease-modifying therapies; thus, symptom management remains the primary treatment for patients. The complex etiologies and pathophysiological mechanisms of RAP and CP pose challenges in developing therapies, necessitating suitable animal models for preclinical drug screening to successfully translate into clinical trials. The goal of this review is to provide information on relevant preclinical models that can be used for drug screening.

Methodology

Publications reporting on animal models in pancreatitis research were sourced from medical and academic journals. While there were many possible models for pancreatitis research identified, many lacked clinical relevance and/or feasibility. As such, we determined that it was necessary to develop a set of criteria to determine a model's relevance to preclinical drug screening. The overarching criteria we used were based on whether the model produced a phenotype that recapitulates RAP or CP in humans and on whether the model was toxic or had negative side effects in the animals. We also considered the model's feasibility and relevance to researchers. Models were classified into either the 'relevant' or 'non-relevant' categories. Models

published prior to 2013 that were referenced in at least one other publication in the past 10 years and models published prior to 2018 that were referenced in at least one other publication in the past five years were considered "relevant". Models published after 2018 were categorized based on the model's feasibility and replicability through the results of its respective study. Conversely, models with limited evidence, severe adverse effects, limited feasibility due to ethical and financial concerns, and those with limited clinical relevance were considered "non-relevant". Models that have been previously used in preclinical drug screening research were also classified as "relevant".

Figure 1: Evaluation of Preclinical models



Results

74 articles were analyzed for preclinical models used for pancreatitis, there were a total of 48 different preclinical models identified. Of these, 19 were classified as 'relevant' and 29 were classified as 'non-relevant' for preclinical drug screening (Figure 1). Relevant models according to the established criteria are listed in Table 1. A full list of all models reviewed is available on our website. Drugs that have been tested in specific models are also listed (Table 1). Dabigatran, a type of blood thinner, was tested in trypsin dependent GEMM models. Pirfenidone, an antifibrotic drug, was tested in two different models, L-arginine, a type of amino acid model, and in the cerulein model. Proglumide, a cholecystokinin (CCK) antagonist drug, and Baicalin, a flavonoid glycoside, were only tested in the cerulein model. Adipose stem cell therapy¹ and FGF21² therapy were tested in the ETOH and cerulein model. The Orai inhibitor, CM5480 was tested in the cerulein model.

Discussion

We have compiled a comprehensive list of preclinical models which may be used for pancreatitis research and listed those that are more relevant based on use and feasibility. While not all the relevant models have been utilized for preclinical drug screening, each holds the potential for such applications. The relevance of these models varies depending on the specific use. Several of the preclinical models have already been used in drug testing for pancreatitis including dabigatran, pirfenidone, proglumide, baicalin, adipose stems cells, FGF21 therapy, and CM5480, an Orail inhibitor.

We provide this information to expedite drug screening studies in relevant preclinical models, thereby accelerating the development of therapies for this therapy this disease.

Table 1: List of Relevant Preclinical Models (chronic pancreatitis)

Model	Drug Testing*	
GEMM Models		
PRSS1 R122H ³	Dabigatran	
transgenic PRSS1R122H ⁴	Dabigatran	
The T7D23A mouse model ⁵	Dabigatran	
T7K24R mouse model ⁶	Dabigatran	
hPRSS1 R122H /N291 ⁷		
PRRS2 and PRSS1 R122H ⁸		
Atg5-knockout /deletion ^{9-12 13}		
Atg7-knockout /deletion ^{14,15}		
IKKa knockout /deletion ¹⁵⁻¹⁷		
PNLIPP p.T122M ¹⁸		
Chemical Models (bile)		
TNBS (intraductal) ^{17,19,20}		
Amino Acid Models		
L-arginine ^{15,17,21-26}	Pirfenidone	
Nongenic Mouse Models		
Pancreatic duct ligation ^{27,28}		
Secretagogue Models		
Cerulein ^{15 1,2,17}	Proglumide, Pirfenidone, Baicalin, adipose stem cell, FGF21	

Mechanical	
Ligation/Obstruction ^{15,17,22,28-42}	
Alcohol induced environmental modulators	
ETOH +HFD chronic (10 week) ^{2,15,17,43}	Adipose stem cell, FGF21

*not complete list.

References

1. Sun Z, Gou W, Kim DS, et al. Adipose Stem Cell Therapy Mitigates Chronic Pancreatitis via Differentiation into Acinar-like Cells in Mice. *Mol Ther*. Nov 1 2017;25(11):2490-2501. doi:10.1016/j.ymthe.2017.06.016

2. Hernandez G, Luo T, Javed TA, et al. Pancreatitis is an FGF21-deficient state that is corrected by replacement therapy. *Sci Transl Med.* Jan 8 2020;12(525)doi:10.1126/scitranslmed.aay5186

3. Huang H, Swidnicka-Siergiejko AK, Daniluk J, et al. Transgenic Expression of PRSS1(R122H) Sensitizes Mice to Pancreatitis. *Gastroenterology*. Mar 2020;158(4):1072-1082.e7. doi:10.1053/j.gastro.2019.08.016

4. Gui F, Zhang Y, Wan J, et al. Trypsin activity governs increased susceptibility to pancreatitis in mice expressing human PRSS1R122H. *J Clin Invest*. Jan 2 2020;130(1):189–202. doi:10.1172/jci130172

5. Geisz A, Sahin-Tóth M. A preclinical model of chronic pancreatitis driven by trypsinogen autoactivation. *Nat Commun*. Nov 28 2018;9(1):5033. doi:10.1038/s41467-018-07347-y

6. Jancsó Z, Sahin-Tóth M. Mutation That Promotes Activation of Trypsinogen Increases Severity of Secretagogue-Induced Pancreatitis in Mice. *Gastroenterology*. Mar 2020;158(4):1083-1094. doi:10.1053/j.gastro.2019.11.020

7. Athwal T, Huang W, Mukherjee R, et al. Expression of human cationic trypsinogen (PRSS1) in murine acinar cells promotes pancreatitis and apoptotic cell death. *Cell Death Dis*. Apr 10 2014;5(4):e1165. doi:10.1038/cddis.2014.120

8. Wang J, Wan J, Wang L, Pandol SJ, Bi Y, Ji B. Wild-Type Human PRSS2 and PRSS1(R122H) Cooperatively Initiate Spontaneous Hereditary Pancreatitis in Transgenic Mice. *Gastroenterology*. Jul 2022;163(1):313-315.e4. doi:10.1053/j.gastro.2022.03.009

9. Diakopoulos KN, Lesina M, Wörmann S, et al. Impaired autophagy induces chronic atrophic pancreatitis in mice via sex- and nutrition-dependent processes. *Gastroenterology*. Mar 2015;148(3):626-638.e17. doi:10.1053/j.gastro.2014.12.003

10. Gukovsky I, Gukovskaya AS. Impaired autophagy triggers chronic pancreatitis: lessons from pancreas-specific atg5 knockout mice. *Gastroenterology*. Mar 2015;148(3):501-5. doi:10.1053/j.gastro.2015.01.012

 Ichimura Y, Komatsu M. Pathophysiological Role of Autophagy: Lesson from Autophagy-Deficient Mouse Models. *Experimental Animals*. 2011;60(4):329-345. doi:10.1538/expanim.60.329

12. Mareninova OA, Hermann K, French SW, et al. Impaired autophagic flux mediates acinar cell vacuole formation and trypsinogen activation in rodent models of acute pancreatitis. *J Clin Invest*. Nov 2009;119(11):3340-55. doi:10.1172/jci38674

13. Hashimoto D, Ohmuraya M, Hirota M, et al. Involvement of autophagy in trypsinogen activation within the pancreatic acinar cells. *J Cell Biol*. Jun 30 2008;181(7):1065-72. doi:10.1083/jcb.200712156

14. Antonucci L, Fagman JB, Kim JY, et al. Basal autophagy maintains pancreatic acinar cell homeostasis and protein synthesis and prevents ER stress. *Proc Natl Acad Sci U S A*. Nov 10 2015;112(45):E6166-74. doi:10.1073/pnas.1519384112

15. Klauss S, Schorn S, Teller S, et al. Genetically induced vs. classical animal models of chronic pancreatitis: a critical comparison. *The FASEB Journal*. 2018;32(11):5778-5792. doi:<u>https://doi.org/10.1096/fj.201800241RR</u>

16. Li N, Wu X, Holzer RG, et al. Loss of acinar cell IKKα triggers spontaneous pancreatitis in mice. *J Clin Invest*. May 2013;123(5):2231-43. doi:10.1172/jci64498

17. Saloman JL, Albers KM, Cruz-Monserrate Z, et al. Animal Models: Challenges and Opportunities to Determine Optimal Experimental Models of

Pancreatitis and Pancreatic Cancer. *Pancreas*. Jul 2019;48(6):759-779. doi:10.1097/mpa.00000000001335

18. Zhu G, Wilhelm SJ, George LG, et al. Preclinical mouse model of a misfolded PNLIP variant develops chronic pancreatitis. *Gut*. Jul 2023;72(7):1340-1354. doi:10.1136/gutjnl-2022-327960

19. Zhu Y, Mehta K, Li C, et al. Systemic administration of anti-NGF increases A-type potassium currents and decreases pancreatic nociceptor excitability in a rat model of chronic pancreatitis. *Am J Physiol Gastrointest Liver Physiol*. Jan 1 2012;302(1):G176-81. doi:10.1152/ajpgi.00053.2011

20. Xu GY, Winston JH, Shenoy M, Yin H, Pasricha PJ. Enhanced excitability and suppression of A-type K+ current of pancreas-specific afferent neurons in a rat model of chronic pancreatitis. *Am J Physiol Gastrointest Liver Physiol*. Sep 2006;291(3):G424-31. doi:10.1152/ajpgi.00560.2005

21. Mizunuma T, Kawamura S, Kishino Y. Effects of injecting excess arginine on rat pancreas. *J Nutr*. Mar 1984;114(3):467-71. doi:10.1093/jn/114.3.467

22. Zhang J, Rouse RL. Histopathology and pathogenesis of caerulein-, duct ligation-, and arginine-induced acute pancreatitis in Sprague-Dawley rats and C57BL6 mice. *Histol Histopathol*. Sep 2014;29(9):1135-52. doi:10.14670/hh-29.1135

23. Wang Y, Kayoumu A, Lu G, et al. Experimental Models in Syrian Golden Hamster Replicate Human Acute Pancreatitis. *Sci Rep*. Jun 15 2016;6:28014. doi:10.1038/srep28014

24. Toma H, Winston J, Micci MA, Shenoy M, Pasricha PJ. Nerve growth factor expression is up-regulated in the rat model of L-arginine-induced acute pancreatitis. *Gastroenterology*. Nov 2000;119(5):1373-81. doi:10.1053/gast.2000.19264

25. Dawra R, Sharif R, Phillips P, Dudeja V, Dhaulakhandi D, Saluja AK. Development of a new mouse model of acute pancreatitis induced by administration of L-arginine. *Am J Physiol Gastrointest Liver Physiol*. Apr 2007;292(4):G1009-18. doi:10.1152/ajpgi.00167.2006

26. Delaney CP, McGeeney KF, Dervan P, Fitzpatrick JM. Pancreatic atrophy: a new model using serial intra-peritoneal injections of L-arginine. *Scand J Gastroenterol*. Dec 1993;28(12):1086-90. doi:10.3109/00365529309098314

27. Peng C, Tu G, Yu L, et al. Murine Chronic Pancreatitis Model Induced by Partial Ligation of the Pancreatic Duct Encapsulates the Profile of Macrophage in Human Chronic Pancreatitis. *Front Immunol*. 2022;13:840887. doi:10.3389/fimmu.2022.840887

28. Sendler M, Beyer G, Mahajan UM, et al. Complement Component 5 Mediates Development of Fibrosis, via Activation of Stellate Cells, in 2 Mouse Models of Chronic Pancreatitis. *Gastroenterology*. Sep 2015;149(3):765-76.e10. doi:10.1053/j.gastro.2015.05.012

29. Kaiser AM, Saluja AK, Sengupta A, Saluja M, Steer ML. Relationship between severity, necrosis, and apoptosis in five models of experimental acute pancreatitis. *Am J Physiol*. Nov 1995;269(5 Pt 1):C1295-304. doi:10.1152/ajpcell.1995.269.5.C1295

30. Mooren F, Hlouschek V, Finkes T, et al. Early changes in pancreatic acinar cell calcium signaling after pancreatic duct obstruction. *J Biol Chem*. Mar 14 2003;278(11):9361-9. doi:10.1074/jbc.M207454200

31. Watanabe S, Abe K, Anbo Y, Katoh H. Changes in the mouse exocrine pancreas after pancreatic duct ligation: a qualitative and quantitative histological study. *Arch Histol Cytol*. Aug 1995;58(3):365-74. doi:10.1679/aohc.58.365

32. Ohshio G, Saluja A, Steer ML. Effects of short-term pancreatic duct obstruction in rats. *Gastroenterology*. Jan 1991;100(1):196-202. doi:10.1016/0016-5085(91)90601-g

33. Boerma D, Straatsburg IH, Offerhaus GJ, Gouma DJ, van Gulik TM. Experimental model of obstructive, chronic pancreatitis in pigs. *Dig Surg*. 2003;20(6):520-6. doi:10.1159/000073688

34. Churg A, Richter WR. Early changes in the exocrine pancreas of the dog and rat after ligation of the pancreatic duct. A light and electron microscopic study. *Am J Pathol.* Jun 1971;63(3):521-46.

35. Isaksson G, Lundquist I, Ihse I. Effects on the exocrine and endocrine pancreas of duct occlusion with two different tissue glues in the rat. *Eur Surg Res.* 1983;15(3):136-44. doi:10.1159/000128345

36. Simpson KW, Batt RM, McLean L, Morton DB. Circulating concentrations of trypsin-like immunoreactivity and activities of lipase and amylase after pancreatic duct ligation in dogs. *Am J Vet Res.* May 1989;50(5):629-32.

37. Tanaka T, Ichiba Y, Fujii Y, Itoh H, Kodama O, Dohi K. New canine model of chronic pancreatitis due to chronic ischemia with incomplete pancreatic duct obstruction. *Digestion*. 1988;41(3):149-55. doi:10.1159/000199767

38. Vigna SR, Shahid RA, Nathan JD, McVey DC, Liddle RA. Leukotriene B4 mediates inflammation via TRPV1 in duct obstruction-induced pancreatitis in rats. *Pancreas*. Jul 2011;40(5):708-14. doi:10.1097/MPA.0b013e318214c8df
39. Patel AG, Reber PU, Toyama MT, Ashley SW, Reber HA. Effect of pancreaticojejunostomy on fibrosis, pancreatic blood flow, and interstitial pH in chronic pancreatitis: a feline model. *Ann Surg*. Nov 1999;230(5):672-9. doi:10.1097/00000658-199911000-00009

40. Reber PU, Patel AG, Lewis MP, Ashley SW, Reber HA. Stenting does not decompress the pancreatic duct as effectively as surgery in experimental chronic pancreatitis. *Surgery*. Sep 1998;124(3):561-7.

41. Zhang TT, Wang L, Wang DB, Huang ZJ, Li YH, Lu JP. Correlation between secretin-enhanced MRCP findings and histopathologic severity of chronic pancreatitis in a cat model. *Pancreatology*. Sep-Oct 2013;13(5):491-7. doi:10.1016/j.pan.2013.08.003

42. Sakakibara A, Okumura N, Hayakawa T, Kanzaki M. Ultrastructural changes in the exocrine pancreas of experimental pancreatolithiasis in dogs. *Am J Gastroenterol*. Jul 1982;77(7):498-503.

43. McIlwrath SL, Westlund KN. Pharmacological attenuation of chronic alcoholic pancreatitis induced hypersensitivity in rats. *World J Gastroenterol*. Jan 21 2015;21(3):836-53. doi:10.3748/wjg.v21.i3.836